

Typing single polymorphic nucleotides in mitochondrial DNA as a way to access Middle Pleistocene DNA

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In this study, we have used a technique designed to target short fragments containing informative mitochondrial substitutions to extend the temporal limits of DNA recovery and study the molecular phylogeny of *Ursus deningeri*. We present a cladistic analysis using DNA recovered from 400 kyr old *U. deningeri* remains, which demonstrates *U. deningeri*'s relation to *Ursus spelaeus*. This study extends the limits of recovery from skeletal remains by almost 300 kyr. Plant material from permafrost environments has yielded DNA of this age in earlier studies, and our data suggest that DNA in teeth from cave environments may be equally well preserved.

Keywords: evolution; Ursidae; ancient DNA; Atapuerca

1. INTRODUCTION

When DNA from prehistoric tissue was introduced as a genetic source material to the scientific community, a possibility to study evolutionary processes seemed realistic. Attempts were also made to extract DNA from geological material (Cano *et al.* 1993; Woodward *et al.* 1994), and contemporary authentication criteria seemed to support some of these results (Bada *et al.* 1994). However, when higher stringency was applied to these types of study, it became evident that several of the oldest DNA sequences were based on contaminating DNA (Zischler *et al.* 1995;

Austin *et al.* 1997). Most ancient DNA sequences that have been thoroughly authenticated are less than 100 000 years, with only a few exceptions (Loreille *et al.* 2001; Willerslev *et al.* 2003). The number of copies should increase with decreased fragment size, as DNA in ancient tissue is degraded (Smith *et al.* 2003; Noonan *et al.* 2005). Therefore, it may be possible to extend the age limit of amplifiable DNA by targeting short fragments. Further, by selecting specific informative point mutations, it may be possible to apply evolutionary questions to DNA from fossils of great age. We investigate this possibility here by extracting and amplifying specific polymorphic sites in cytochrome *b* from bear material (*Ursus deningeri*) excavated from the Atapuerca cave complex in northern Spain, a site from which younger prehistoric material has yielded DNA (Anderung *et al.* 2005).

Ursus deningeri was a part of the Pleistocene European Ursidae family represented by two main evolutionary lineages, one resulting in the extant brown bear (*Ursus arctos*) and the other leading to the cave bear (*Ursus spelaeus*). These two lineages diverged in Eurasia about 1.2 million years ago (Loreille *et al.* 2001), and their common ancestor has been suggested to be the so-called Etruscan bear, *Ursus etruscus* (Soergel 1912). *Ursus dolinensis* has been recently suggested to represent the earliest member of the speleoid lineage (García & Arsuaga 2001). *Ursus deningeri* is first recognized in the European fossil record at around 600 kyr ago, and lasts until the Late Middle Pleistocene (García *et al.* 1997) when *U. spelaeus* appears. This latter species went extinct at approximately 12 kyr (BP) (Kurtén 1968). Cave bears are thus believed to have evolved as a single, unbroken lineage *U. dolinensis*–*U. deningeri*–*U. spelaeus* (García & Arsuaga 2001). To investigate the possibility of using molecular data from Middle Pleistocene mammals, we have focused on the genetic relationship between *U. deningeri* and *U. spelaeus*.

2. MATERIAL AND METHODS

A total of 25 bear samples from five different well-studied Pleistocene fossil sites in Europe were analysed (figure 1). The *U. deningeri* samples ($n=5$) were recovered from the Sima de los Huesos site in the Sierra de Atapuerca (García *et al.* 1997), and date to approximately 400 kyr (Bischoff *et al.* 2003). The site is located at a depth of 25 m below the present ground surface, with conditions of constant humidity and temperature (11 °C). The *U. spelaeus* samples analysed were recovered from: (i) Late Middle Pleistocene (ca 230 kyr old) or Early Upper Pleistocene (ca 120 kyr old) travertines of Ehringsdorf (Weimar), Central Germany ($n=1$; Kurtén 1975; Kahlke *et al.* 2002); (ii) 30-kyr-old sediments of Vindija cave, Croatia ($n=6$; Smith *et al.* 1999); and (iii) the undated Upper Pleistocene site of Coro Tracito cave, Pyrenees, Spain ($n=3$; Torres *et al.* 1998). In addition, a set of *U. arctos* samples, originating from the Ehringsdorf (Weimar) travertine sequence (from ca 220–110 kyr) and the last interglacial travertine sands of Taubach (Weimar; $n=10$; Kurtén 1977) of around 120 kyr, was analysed.

DNA in prehistoric material is present in minute amounts and highly fragmented (Hofreiter *et al.* 2001). Since the probability of amplifying short fragments of DNA is likely to be higher than the amplifying longer fragments (Smith *et al.* 2003; Noonan *et al.* 2005), genetic markers were selected to be short and specific for bear mitochondrial polymorphisms from the cytochrome *b* gene that could differentiate among three different species of ursids: *U. arctos*, *U. spelaeus* and *U. americanus*. As we aimed for minimal amplicon size, primer sites were always less than 6 bp upstream and downstream from the polymorphic site. DNA was extracted from the material with a phosphate buffer, and purified and concentrated with a hybridization/magnetic separation technique (Anderung *et al.*

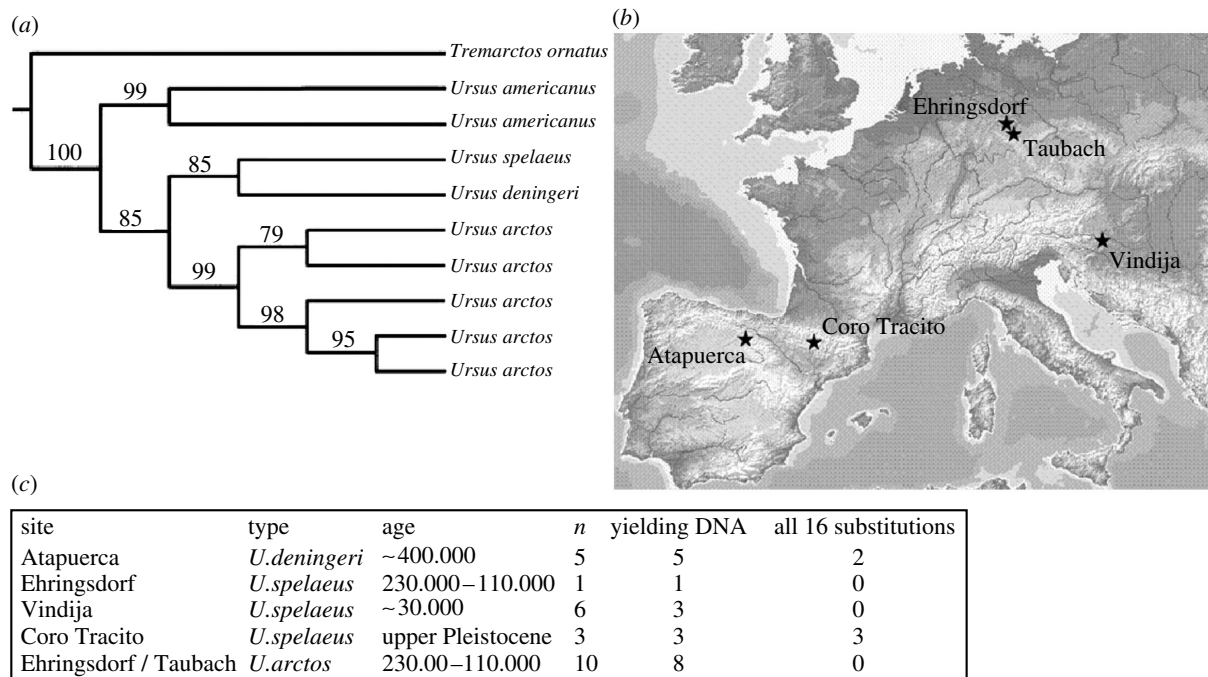


Figure 1. (a) Phylogenetic position of *Ursus deningeri*. Maximum parsimony tree using molecular data from cytochrome *b*. The grouping of *U. deningeri* and *U. spelaeus* was well supported (1000 replicates). (b) Sites for the caves with bear samples. (c) Out of 25 samples, 20 yielded bear DNA, out of which 5 yielded all 16 polymorphic sites.

2005). The DNA was then amplified and typed with pyrosequencing technology.

3. RESULTS

A total of 20 individuals yielded DNA (see electronic supplementary material) and five of them yielded all the 16-targeted polymorphisms in cytochrome *b*. From two of the oldest samples, 400 kyr old *U. deningeri* (Bischoff *et al.* 2003), we were able to pyrosequence all 16 polymorphisms (the remaining three yielded DNA, but not all targeted sites). Seven of the *U. spelaeus* samples yielded DNA, three of them yielded the same information from cytochrome *b* as *U. deningeri* or more. For the Ehringsdorf and Taubach *U. arctos* samples, we were able to pyrosequence up to 13 polymorphic sites. When the genetic data were compared with previously published *Ursus* sp. sequences, the phylogenetic tree gave strong support for the placement of *U. deningeri* together with *U. spelaeus* (figure 1).

4. DISCUSSION

Owing to the non-overlapping chronological ranges of *U. deningeri* and *U. spelaeus*, there are two possible hypotheses regarding the evolutionary relationship between them. First, they could be sister species, that diverged about 600 kyr ago. Second, *U. deningeri* could be the direct ancestor of *U. spelaeus*. Obtaining DNA sequences from both species using samples collected close to the transition period should allow these two hypotheses to be tested. Although *U. deningeri* and *U. spelaeus* had identical DNA sequences, it is likely that the total sequence length (41 bp) obtained in this study is insufficient to resolve these two hypotheses. Additional sequence data

including more polymorphic sites, e.g. variable within *U. spelaeus*, would be needed.

Authentication followed the standard rules (Cooper & Poinar 2000). The work was carried out in a new ancient DNA laboratory, where no DNA work (ancient or modern) had been undertaken. Replication was carried out in an ancient DNA laboratory, where there had been no previous work with bear DNA. When the replication was carried out, ancient samples of elephant (*Loxodonta africana*) and lemming (*Lemmus sibiricus*) were included as negative controls. These did not produce bear sequences, but did give appropriate results when targeted with appropriate species-specific primers. Further, the *U. deningeri* samples did not produce amplicons with the elephant- and lemming-specific primers. A non-random degradation pattern has been suggested to be a problem in ancient DNA (Gilbert *et al.* 2003; Banerjee & Brown 2004). The similarity between recent and the *ca* 220–110 kyr old *U. arctos* in this study indicates that the sequences obtained from *U. deningeri* are not non-randomly degraded *U. arctos* fragments. Since the minimal fragment sizes targeted could not be efficiently quantified with real-time PCR (real-time PCR techniques demand larger fragments), we based sequences and reactions on a minimum of three extractions (with the exception for some of the younger *U. arctos* that were only extracted twice).

Targeting short DNA fragments appears to be a powerful tool when working with highly degraded DNA. Using this technique, we were able to retrieve genetic information from 400 kyr old *U. deningeri* samples. Typing polymorphic sites in short mitochondrial fragments may yield access to genetic data

on other Middle Pleistocene animals in a similar fashion.

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- Anderung, C. *et al.* 2005 Prehistoric contacts over the Straits of Gibraltar indicated by genetic analysis of Iberian Bronze Age cattle. *Proc. Natl Acad. Sci. USA* **102**, 8431–8435. (doi:10.1073/pnas.0503396102)
- Austin, J. J., Ross, A. J., Smith, A. B., Fortey, R. A. & Thomas, R. H. 1997 Problems of reproducibility—does geologically ancient DNA survive in amber-preserved insects? *Proc. R. Soc. B* **264**, 467–474. (doi:10.1098/rspb.1997.0067)
- Bada, J. L., Wang, X. S., Poinar, H. N., Paabo, S. & Poinar, G. O. 1994 Amino acid racemization in amber-entombed insects: implications for DNA preservation. *Geochim. Cosmochim. Acta* **58**, 3131–3135. (doi:10.1016/0016-7037(94)90185-6)
- Banerjee, M. & Brown, T. 2004 Non-random DNA damage resulting from heat treatment: implications for sequence analysis of ancient DNA. *J. Archaeol. Sci.* **31**, 59–63. (doi:10.1016/S0305-4403(03)00099-2)
- Bischoff, J. L., Shamp, D. D., Aramburu, A., Arsuaga, J. L., Carbonell, E. & Bermudez de Castro, J. M. 2003 The Sima de los Huesos hominids date to beyond U/Th equilibrium (>350 kyr) and perhaps to 400–500 kyr: new radiometric dates. *J. Archaeol. Sci.* **30**, 275–280. (doi:10.1006/jasc.2002.0834)
- Cano, R. J., Poinar, H. N., Pieniazek, N. J., Acra, A. & Poinar Jr, G. O. 1993 Amplification and sequencing of DNA from a 120–135-million-year-old weevil. *Nature* **363**, 536–538. (doi:10.1038/363536a0)
- Cooper, A. & Poinar, H. N. 2000 Ancient DNA: do it right or not at all. *Science* **289**, 1139. (doi:10.1126/science.289.5482.1139b)
- García, N. & Arsuaga, J. L. 2001 *Ursus dolinensis*: a new species of Early Pleistocene ursid from Trincheras Dolina, Atapuerca (Spain). *Comptes Rendus de l'Académie des Sciences de Paris, série II* **332**, 717–725.
- García, N., Arsuaga, J. L. & Torres, T. 1997 The carnivore remains from the Sima de los Huesos Middle Pleistocene site (Sierra de Atapuerca, Spain). *J. Hum. Evol.* **33**, 155–174.
- Gilbert, M. T., Willerslev, E., Hansen, A. J., Barnes, I., Rudbeck, L., Lynnerup, N. & Cooper, A. 2003 Distribution patterns of postmortem damage in human mitochondrial DNA. *Am. J. Hum. Genet.* **72**, 32–47. (doi:10.1086/345378)
- Hofreiter, M., Serre, D., Poinar, H. N., Kuch, M. & Paabo, S. 2001 Ancient DNA. *Nat. Rev. Genet.* **2**, 353–359. (doi:10.1038/35072071)
- Kahlke, R.-D., Maul, L. C., Meyrick, R. A., Stebich, M. & Grasselt, T. 2002 The quaternary sequence from the late Middle to Upper Pleistocene site of Weimar-Ehringsdorf. In *The quaternary of central Germany (Thuringia and surroundings)* (ed. R. A. Meyrick & D. C. Schereve), pp. 163–177. London, UK: Quaternary Research Association.
- Kurtén, B. 1968 *Pleistocene mammals of Europe*. London, UK: Weidenfeld & Nicolson.
- Kurtén, B. 1975 Fossile Reste von Hyänen und Bären (Carnivora) aus den Travertinen von Weimar-Ehringsdorf. In *Das Pleistozän von Weimar-Ehringsdorf. Teil 2* (ed. H.-D. Kahlke) *Abh. Zentr. Geol. Inst., Paläont. Abh.*, vol. 23, pp. 465–484. Berlin, Germany: Akademie-Verlag.
- Kurtén, B. 1977 Bären—und Hyänenreste aus dem Pleistozän von Taubach. In *Das Pleistozän von Taubach bei Weimar* (ed. H.-D. Kahlke) *Quartärpaläontologie*, vol. 2, pp. 361–378. Berlin, Germany: Akademie-Verlag.
- Loreille, O., Orlando, L., Patou-Mathis, M., Philippe, M., Taberlet, P. & Hanni, C. 2001 Ancient DNA analysis reveals divergence of the cave bear, *Ursus spelaeus*, and brown bear, *Ursus arctos*, lineages. *Curr. Biol.* **11**, 200–203. (doi:10.1016/S0960-9822(01)00046-X)
- Noonan, J. P. *et al.* 2005 Genomic sequencing of Pleistocene cave bears. *Science* **309**, 597–599. (doi:10.1126/science.1113485)
- Smith, F. H., Trinkaus, E., Pettitt, P. B., Karavanic, I. & Paunovic, M. 1999 Direct radiocarbon dates for Vindija G1 and Velika Pecina Late Pleistocene hominid remains. *Proc. Natl Acad. Sci. USA* **96**, 12 281–12 286. (doi:10.1073/pnas.96.22.12281)
- Smith, C. I., Chamberlain, A. T., Riley, M. S., Stringer, C. & Collins, M. J. 2003 The thermal history of human fossils and the likelihood of successful DNA amplification. *J. Hum. Evol.* **45**, 203–217. (doi:10.1016/S0047-2484(03)00106-4)
- Soergel, W. 1912 *Das Aussterben diluvialer Säugetiere und die Jagd des diluvialen Menschen*. Jena, Germany: Gustav Fischer.
- Torres, T., Canudo, J. I., Cobo, R. & Cuenca, G. 1998 Cueva Coro Tracito (Tella Sin, Huesca) el primer yacimiento de alta montaña Español de *Ursus spelaeus* Ros.-Hein. Nota Preliminar. *Geogaceta* **24**, 303–306.
- Willerslev, E. *et al.* 2003 Diverse plant and animal genetic records from Holocene and Pleistocene sediments. *Science* **300**, 791–795. (doi:10.1126/science.1084114)
- Woodward, S. R., Weyand, N. J. & Bunnell, M. 1994 DNA sequence from Cretaceous period bone fragments. *Science* **266**, 1229–1232.
- Zischler, H., Hoss, M., Handt, O., von Haeseler, A., van der Kuyl, A. C. & Goudsmit, J. 1995 Detecting dinosaur DNA. *Science* **268**, 1192–1193.